A PRACTICAL SYNTHESIS OF 2-ACETAMIDO-2-DEOXY-3,4-DI-*O-β*-D-GALACTOPYRANOSYL-D-GALACTOPYRANOSE*

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ABSTRACT

Two different sugar derivatives having free hydroxyl groups have been employed for synthesis of the title trisaccharide. In one attempt, benzyl 2-acetamido-6-O-benzyl-2-deoxy- α -D-galactopyranoside (8) was treated with an excess of 2,3,4,6-tetra-O-acetyl- α -D-galactopyranosyl bromide (11), to give a mixture of products which, on fractionation, afforded benzyl 2-acetamido-6-O-benzyl-3,4-di-O-(2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl)- α -D-galactopyranoside (15) in 21% yield. However, in another, preferable approach, benzyl 2-acetamido-6-O-benzyl-2-deoxy-3-O-(2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl)- α -D-galactopyranoside (13) was treated with 11, to produce 15 in 69% yield. Both 8 and 13 were conveniently prepared *via* reductive ring-opening of the respective 4,6-benzylidene acetals. O-Deacetylation of 15, followed by hydrogenolysis, provided the title trisaccharide. The structure of the final product, and of various other intermediates, was established by 1 H- and 13 C-n.m.r. spectroscopy.

INTRODUCTION

The carbohydrate sequence β -Gal-(1 \rightarrow 4)-GalNAc

3 ↑ 1 β-Gal

has been found to be a part of the carbohydrate moiety of the component of T-antigens². The title trisaccharide is also reported to be a part of the core structure of the carbohydrate sequence of the blood-group, M-specific glycopentapeptide, which is further linked to neuraminic acid groups as follows².

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$$\alpha$$
-NeuAc-(2 \rightarrow 3)- β -Gal-(1 \rightarrow 3)
$$\alpha$$
-GalNAc-(1 \rightarrow 3)-Ser
$$\alpha$$
-NeuAc-(2 \rightarrow 3)- β -Gal-(1 \rightarrow 4)

We had already accomplished the synthesis of 2-acetamido-2-deoxy-4-O- β -D-galactopyranosyl-D-galactose³, and also reported that synthetic phenyl 2-acetamido-2-deoxy-3-O- β -D-galactopyranosyl- α -D-galactopyranoside acts as an acceptor for one of the sialyltransferases present in human serum⁴. The possibility of glycosyltransferases as tumor markers has been demonstrated by various investigators⁵. The title trisaccharide was needed in our laboratory for specificity testing of human sialyltransferases, and we now describe a practical and elegant synthesis of it.

RESULTS AND DISCUSSION

According to Bovin *et al.*⁶, reaction of benzyl 2-acetamido-6-O-acetyl-2-deoxy- α -D-glucopyranoside with an equimolar proportion of 2,3,4,6-tetra-O-acetyl- α -D-galactopyranosyl bromide afforded the corresponding 3-O-substituted disaccharide derivative. In a recent review, Paulsen⁷ mentioned that the title trisaccharide had been α -linked to CH₂CH₂NHCO(CH₂)₄CO₂Me in his laboratory, and that the order in which the two D-galactosyl groups are attached to the GalNAc residue is important. The protected, disubstituted trisaccharide is obtained in high yield if the first galactosyl group is coupled at OH-3, and the second at OH-4.

Consequently, based on these two observations $^{6.7}$, we anticipated in our first approach that the reaction of an appropriately protected 2-acetamido-2-deoxy-D-galactose (having both OH-3 and -4 free) with an excess of compound 11 would afford the expected trisaccharide derivative. It is also apparent that the desired 6-O-substituted derivative, e.g., benzyl 2-acetamido-6-O-benzyl(or 6-O-acetyl)-2-deoxy- α -D-galactopyranoside, can be obtained from benzyl 2-acetamido-2-deoxy- α -D-galactopyranoside in three steps, namely, acetonation to give the 3,4-O-isopropylidene derivative as the main product, protection of the primary hydroxyl group thereof by benzylation or acetylation, and O-deisopropylidenation.

Very recently, Garegg and Hultberg⁸ reported a novel method of reductive ring-opening of carbohydrate benzylidene acetals with sodium cyanoborohydride in HCl-ether. Under these conditions, benzyl 2-acetamido-4,6-O-benzylidene-2-deoxy- α -D-galactopyranoside (2), preparable from 1, gave 8 in 56 $^{\circ}c$ yield. It may also be pointed out that pure benzyl 2-acetamido-2-deoxy- α -D-galactopyranoside is obtained by removal of the 4,6-O-benzylidene group from 2, as it has been reported⁹ that, on treatment with benzyl alcohol containing a catalytic amount of dry HCl, commercially available 2-acetamido-2-deoxy-D-galactose gives a mixture of anomers that are conveniently separated by conversion into the 4,6-O-benzylidene

TABLE I
³ C-N M R ^a CHEMICAL SHIFTS ^b

Atom	Compound						
	1	7	8	9	10		
C-1	96.08	96.36	96.30	96.96	96.31		
C-2	49.59	48.28	49.57	48.58	50.09		
C-3	67 19	75.89	68.41	76.17	68.09		
C-4	67.55	64.36	67.07	65 23	77.03		
C-5	71.34	71 44	69.64	70.42	74.30		
C-6	60.53	60.58	72.08	72.61	72.16		
COCH ₃	22.49	22.63	22.52	23.00	22 56		
CH ₂ Ph	67.99	67.87	67.91	68 54	67.56		
-		69.88	69.55	70.03	68.91		
					69.14		
C=O	169.22	169.09	169.30	169.53	169 34		

^aAt 25.2 MHz; solution in Me₂SO-d₆. ^bIn p.p.m. downfield from Me₄Si (internal).

acetal followed by fractional recrystallization. In other words, the reductive-cleavage technique provides a rapid method for preparation of the diol **8**, required for further glycosylation. The structure of diol **8** was unambiguously supported by its 13 C-n.m.r. spectrum (see Table I). The pronounced, downfield shift of 11.55 p.p.m. exhibited by C-6 on benzylation, and the upfield shift (1.70 p.p.m.) of C-5, confirmed the position of substitution in **8**. In the present studies, we have also prepared benzyl 2-acetamido-3,6-di-O-benzyl- α -D-galactopyranoside³ (**9**) by reductive cleavage of the benzylidene acetal of compound **4**, whereas, in a previous attempt³, compound **9** was obtained by selective benzylation of benzyl 2-acetamido-3-O-benzyl-2-deoxy- α -D-galactopyranoside (**7**) by the phase-transfer-catalysis method.

The reaction of compound 2 with allyl bromide in N,N-dimethylformamide in the presence of barium oxide and barium hydroxide produced crystalline 3 which, on treatment with aqueous acetic acid at 100°, gave 5 in 81% yield. On benzylation, compound 5 afforded 6 which, on O-deallylation with potassium tert-butoxide in dimethyl sulfoxide¹⁰, gave 10 in 76% yield. The structures of the benzylated derivatives 7, 8, 9, and 10 were established by ¹³C-n.m.r. spectroscopy (see Table I).

The coupling reaction of diol 8 with bromide 11 in dichloromethane was conducted in the presence of silver triflate and 1,1,3,3-tetramethylurea¹¹, the reaction being monitored by t.l.c. (4:1 chloroform-acetone) which, after 3 days, showed a major spot corresponding to benzyl 2-acetamido-6-O-benzyl-2-deoxy-3-O-(2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl)- α -D-galactopyranoside (13), trisaccharide derivative 15 as a minor product, and a significant amount of the starting material 8. Additional amounts of the halide and catalyst were introduced, to give 15; nevertheless, a considerable amount of starting material remained under these coupling conditions.

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In another approach, we observed that the reductive cleavage⁸ of benzyl 2-acetamido-4,6-O-benzylidene-2-deoxy-3-O-(2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl)- α -D-galactopyranoside (12) afforded 13, having an O-benzyl group selectively at the primary position. O-Deacetylation of 13 provided benzyl 2-acetamido-6-O-benzyl-2-deoxy-3-O- β -D-galactopyranosyl- α -D-galactopyranoside (14) in 87% yield. The downfield shift of 11.58 p.p.m. exhibited by C-6 on benzylation (see Table II), and the upfield shift of C-5 (1.73 p.p.m.) confirmed the position of substitution in 14. The complete absence of a C-6 signal in the region of 60–63 p.p.m. also confirmed that reductive cleavage of the benzylidene acetal 12 had given only the 6-O-benzyl derivative 13. Treatment of alcohol 13 with bromide 11 in anhydrous dichloromethane in the presence of silver triflate and 1.1.3,3-tetramethylurea¹¹ afforded trisaccharide derivative 15 in 69% yield

It is unclear why, on coupling with bromide 11, diol 8 gives a low yield of expected compound 15, although slow formation of disaccharide derivative 13 was observed. It is possible that, for glycosylation of diol 8, changes in the reaction conditions, e.g., in solvent and catalyst (probably a catalyst which is not hygroscopic) and use of a higher temperature, might provide compound 15 in appreciable yield. Nevertheless, based upon the present experimental observation, the use of "aglycon" hydroxide 13, already having a 3-O-D-galactosyl unit, is strongly preferred for

TABLE II
25.2-MHz, ¹³C-N M R CHEMICAL SHIFTS ^a

Atom	Compound							
	14	16	17α		17β			
C-1	96.46	97.93	92.37		96.20			
C-2	48,34	50.65	50.57		53.99			
C-3	75 53	77.86	77.62		80.64			
C-4	67.58	77.32	76.95		76.05			
C-5	69.61	71.73	70.89		75.25			
C-6	72.11	72.33	61.76		61.53			
C=O	169.48	173.91	175.62		175.94			
CH ₃	22.57	22,77	23.19		23.41			
C-1"	103.55	104.78		104.24				
C-2'	70.64	73.13		71.72				
C-3'	73.24	74.51		73.59				
C-4'	68.05	70.22		69.74				
C-5'	75.21	76.23		75.86				
C-6'	60,42	62,40		62.13				
C-1"		106.35	105.92		106.22			
C-2"		74.20		72.28				
C-3"		74,90		73.72				
C-4"		70.36		69.74				
C-5"		76.77		76.05				
C-6"		62.74		62.20				

[&]quot;In p.p.m. downfield from Me₄Si. The solvent was Me₂SO- d_6 for 14, CD₃OD for 16, and D₂O for 17. The reference (Me₄Si) was internal for solutions in Me₂SO- d_6 and CD₃OD, and external for that in D₂O.

preparation of compound 15. An "aglycon" hydroxide, similar to 13 but having an ester as the protecting group on the primary hydroxyl group of the GalNAc residue, has been recommended for such glycosylation¹²; however, it is apparent that preparation of such a disaccharide "aglycon" hydroxide with a 6-ester group on the GalNAc unit is likely to be accomplished by selective acetylation of the corresponding 4,6-diol derivative. For example, should there be an interest in employing benzyl 2-acetamido-6-O-benzoyl-2-deoxy-3-O-(2,3,4,6-tetra-O-acetyl-β-D-galactopyranosyl)- α -D-galactopyranoside for further glycosylation at O-4 of GalNAc, this compound will have to be obtained by selectively benzoylating O-6 of the diol obtained by removal of the 4,6-O-benzylidene group of 12. On the other hand, reductive cleavage8 of 12 directly afforded, in one step, the desired, key intermediate having a 6-O-benzyl group on the GalNAc residue. The use of reductive cleavage of 4,6-O-benzylidene acetals, recently introduced into the field of carbohydrate chemistry, seems to be excellent for procurement of suitably protected sugars having free alcohol groups. To the best of our knowledge, the present investigation provides the first example of the technique of reductive ring-opening of a 4,6-benzylidene acetal in order to utilize the product successfully in the preparation of ap-

propriately protected disaccharides having a free alcohol group for further synthesis of higher saccharides.

O-Deacetylation¹³ of **15** gave **16**, which, on catalytic hydrogenolysis in glacial acetic acid in the presence of 10°7 Pd–C, produced the title trisaccharide **17** as an amorphous material; its structure was confirmed by ¹³C-n.m.r. spectroscopy.

EXPERIMENTAL

16 P = H

General methods. — Melting points were determined with a Fisher-Johns apparatus and are uncorrected. Optical rotations were measured with a Perkin-Elmer 241 polarimeter at room temperature. Ascending t I.e. was conducted on

plates coated with a 0.25-mm layer of silica gel 60 PF-254 (E. Merck, Darmstadt, Germany); the components were located by exposure to u.v. light, or by spraying the plate with 5% sulfuric acid in ethanol, and heating. Elemental analyses were performed by Robertson Laboratory, Florham Park, New Jersey, U.S.A. N.m.r. spectra were recorded with Varian EM-390 and Varian XL-100 instruments, $^1\text{H-n.m.r.}$ (100 MHz) and $^{13}\text{C-n.m.r.}$ spectra (25.2 MHz) being obtained by use of the Fourier-transform (F.t.) mode, and the positions of the peaks expressed in δ from the signal for tetramethylsilane.

Benzyl 2-acetamido-3-O-allyl-4,6-O-benzylidene-2-deoxy-α-D-galactopy-ranoside (3). — A solution of compound 2 (3.0 g, 7.5 mmol) in N,N-dimethylformamide (50 mL) was stirred for 1 h at room temperature in the presence of barium oxide (5 g), barium hydroxide octahydrate (1.5 g), and allyl bromide (1.25 mL). The resulting, crystalline mass was then poured into cold, 20% acetic acid (100 mL) with stirring; stirring was continued for 15 min, and the white precipitate was filtered off, washed several times with cold water, and recrystallized from hot methanol, to give 3 (2.8 g) in 85% yield; m.p. 250–252°, [α]_D +176.1° (c 1.1, Me₂SO); t.l.c. (5:1 chloroform–acetone): $R_{\rm F}$ 0.8; $\nu_{\rm max}^{\rm KBr}$ 3300 (NH), 1650 (amide), and 700 cm⁻¹ (aromatic).

Anal. Calc. for $C_{25}H_{29}NO_6$: C, 68.32; H, 6.65; N, 3.19. Found: C, 68.14; H, 6.80; N, 3.04.

Benzyl 2-acetamido-3-O-allyl-2-deoxy-α-D-galactopyranoside (5). — A suspension of 3 (2 g) in 60% acetic acid (100 mL) was stirred for 50 min at 100°. Evaporation, followed by several additions and evaporations of water, and then dry toluene, gave a solid mass which was recrystallized from methanol–ether, to give 5 in 81% yield (1.3 g); m.p. 180–181°, $[\alpha]_D$ +182.5° (c 0.6, Me₂SO); t.l.c. (9:1 chloroform–methanol): R_F 0.64; $\nu_{\rm max}^{\rm KBr}$ 3400 (OH), 3300 (NH), 1645 (amide), 730, and 695 cm $^{-1}$ (aromatic); 1 H-n.m.r. data (Me₂SO- d_6): δ 1.84 (s, 3 H, NAc), 4.76 (d, 1 H, $J_{1,2}$ 4 Hz, H-1), 5.06–5.40 (m, 2 H, =CH₂), 5.77–6.15 (m, 1 H, –CH=), 7.40 (m, 5 H, aromatic), and 7.84 (d, 1 H, $J_{\rm NH,2}$ 9 Hz, NH).

Anal. Calc. for $C_{18}H_{25}NO_6$: C, 61.52; H, 7.17; N, 3.99. Found: C, 61.50; H, 7.34; N, 3.86.

Benzyl 2-acetamido-3-O-allyl-4,6-di-O-benzyl-2-deoxy-α-D-galactopyranoside (6). — A solution of 5 (1.3 g, 3.7 mmol) in N,N-dimethylformamide (40 mL) was stirred for 2 days at room temperature in the presence of barium oxide (2.1 g), barium hydroxide octahydrate (0.75 g), and benzyl bromide (2.3 mL, 4 equiv.). After dilution with chloroform (150 mL), 50% acetic acid (50 mL) was added with stirring; stirring was continued for 15 min, and the chloroform layer was separated, washed successively with water, saturated aqueous sodium hydrogen-carbonate, and water, dried (potassium carbonate), and evaporated to dryness. The residue crystallized from ethyl acetate—ether—hexane, to afford 6 in 76% yield (1.5 g); m.p. 155–156°, [α]_D +94.7° (c 1.3, chloroform); t.l.c. (9:1 chloroform—acetone): R_F 0.75; the i.r. spectrum showed the complete absence of hydroxyl group; ¹H-n.m.r. data (Me₂SO-d₆): δ 1.86 (s, 3 H, NAc), 4.78 (d, 1 H, $J_{1,2}$ 4 Hz,

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H-1), 5.08-5.40 (m, 2 H, =CH₂), 5.80-6.16 (m, 1 H, -CH=), and 7.30-7.45 (m, 15 H, aromatic).

Anal. Calc. for $C_{32}H_{37}NO_6$: C, 72.29; H, 7.02; N, 2.64. Found: C, 72.23; H, 7.31; N, 2.61.

Benzyl 2-acetamido-6-O-benzyl-2-deoxy-α-D-galactopyranoside (8). — A mixture of 2 (1.6 g. 4 mmol) and sodium cyanoborohydride (2.262 g. 36 mmol) in dry oxolane (60 mL) containing 3A molecular sieves (8 g), was cooled to 0°. Hydrogen chloride in diethyl ether was added until the solution was acidic (pH paper, gas evolution). After 30 min at 0°, when t.l.c. (3:2 chloroform—acetone) indicated complete reaction, the mixture was poured into ice—water, and extracted with dichloromethane (5 × 40 mL). The combined extracts were successively washed with water, saturated aqueous sodium hydrogencarbonate, and water, dried (anhydrous magnesium sulfate), and evaporated. The solid mass was purified by chromatography on a column of silica gel, with elution with 3:2 chloroform—acetone, to give 8 in 56% yield (900 mg), amorphous; [α]_D +145.1°(c 1.2, Me₂SO); ¹H-n.m.r. data (Me₂SO-d₆): δ 1.93 (s. 3 H. NAc). 4.90 (d. 1 H. J_{1/2} 4 Hz. H-1), and 7.33 (m, 10 H, aromatic).

Anal. Calc. for C₂₂H₂₇NO₆; C, 65.82; H, 6.78; N, 3 49. Found: C, 65.81; H, 6.86; N, 3.29.

Benzyl 2-acetamido-3,6-di-O-benzyl-2-deoxy- α -D-galactopyranoside (9). — Compound 4 (2.445 g, 5 mmol) was transformed into 9 as described for the preparation of 8. The residue was purified by chromatography on a column of silica gel, eluting first with chloroform, then with 9:1 chloroform-acetone, and finally with 5:1 chloroform-acetone, to give 9 in 77% yield (1.9 g), amorphous; $[\alpha]_D$ +121.6° (c 1.4, chloroform), lit. $[\alpha]_D$ +121.5° (c 1, chloroform); tor $[\alpha]_D$ data, see Table I.

Benzyl 2-acetamido-4,6-di-O-benzyl-2-deoxy- α -D-galactopyranoside (10). — A solution of 6 (1.062 g, 2 mmol) and potassium tert-butoxide (1.48 g) in dimethyl sulfoxide (40 mL) was stirred for 3 h at 100° under a nitrogen atmosphere. After being cooled, the mixture was poured into ice-water (100 mL), extracted with chloroform (4 × 50 mL), and the extract washed with water, dried (anhydrous sodium sulfate), and evaporated to dryness. The colored, oily residue in 9:1 (v/v) acetone-water (40 mL) was stirred with vellow mercuric oxide (1 g), a solution of mercuric chloride (900 mg) in 9:1 (v/v) acetone-water (10 mL) was added dropwise, and the mixture was stirred for 3 h, the suspension filtered, and the filtrate evaporated to dryness. The residue was dissolved in chloroform (200 mL), and the solution was successively washed with 10% aqueous potassium iodide and water, dried (anhydrous sodium sulfate), and evaporated to dryness. The residue was purified by chromatography on a column of silica gel, with elution with 9:1 (v.v.) chloroform-acetone, to afford crystalline 10 (750 mg, 76%); m.p. 176–177% (from acetone-ether-hexane), $[\alpha]_D$ +81.3° (c 0.4, Me₂SO); ¹H-n.m.r. data (CDCl₃); δ 1.98 (s. 3 H. NAc), 4.96 (d. 1 H. $J_{1.2}$ 4 Hz, H-1), 5.80 (d. 1 H. J_{NH} 9 Hz, NH), and 7.30–7.50 (m, 15 H, aromatic).

Anal. Calc. for C₂₉H₃₃NO₆: C, 70.85; H, 6.77; N, 2.85. Found: C, 70.75; H, 6.81; N, 2.84.

Benzyl 2-acetamido-4,6-O-benzylidene-2-deoxy-3-O-(2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl)- α -D-galactopyranoside (12). — A solution of 2 (3.45 g, 8.65 mmol) in 1:1 (v/v) benzene-nitromethane (220 mL) was boiled until 60 mL of the solvent had been distilled. The temperature of the solution was adjusted to 40°, and mercuric cyanide (1.88 g) and a solution of bromide 11 (3.55 g, 8.65 mmol) in 1:1 (v/v) benzene-nitromethane (40 mL) were added. The mixture was stirred for 24 h at room temperature, mercuric cyanide (1.2 g) and bromide 11 (1.78 g, 4.33 mmol) were added, and the suspension was stirred for an additional 24 h. The solids were removed by filtration through a Celite pad, and washed with benzene (200 mL). The filtrate and washings were combined, successively washed twice with aqueous potassium iodide solution, saturated aqueous sodium hydrogencarbonate solution, and water, dried (anhydrous sodium sulfate), and evaporated. The residual syrup was purified by chromatography on a column of silica gel, with elution with 6:1 (v/v) chloroform-acetone, to give amorphous 12 in 81% yield (5.12 g); $[\alpha]_D + 103.8^{\circ}$ (c 1.6, Me₂SO); $\nu_{\text{max}}^{\text{KBr}} 3300$ (NH), 1745 (OAc), 1655 (Amide I), 1525 (Amide II), 1220 (OAc), and 700 cm⁻¹ (Ph); ${}^{1}\text{H-n.m.r.}$ data (CDCl₃): δ 1.90, 1.95, 2.0, 2.10 (s each, 15 H, 4 AcO + 1 NAc), 5.50 (s, 1 H, benzylic H), and 7.3-7.6 (m, 10 H, aromatic).

Anal. Calc. for C₃₆H₄₄NO₁₅: C, 59.17; H, 6.07; N, 1.92. Found: C, 59.17; H, 6.09; N, 1.91.

Benzyl 2-acetamido-6-O-benzyl-2-deoxy-3-O-(2,3,4,6-tetra-O-acetyl-β-D-galactopyranosyl)-α-D-galactopyranoside (13). — Compound 13 was prepared from 12 (1.46 g, 2 mmol) as described for 8. The product was purified by chromatography on a column of silica gel, with elution with 6:1 (v/v) chloroformacetone, to give 13 (920 mg, 63%); m.p. 150–152° (ethyl acetate–ether–hexane), $[\alpha]_D$ +66° (c 1.4, chloroform); t.l.c. in 3:1 chloroform–acetone: R_F 0.41; ¹H-n.m.r. data (CDCl₃): δ 1.90, 1.93, 2.0, 2.10 (4 s, 15 H, 4 AcO + NAc) and 7.20–7.40 (m, 10 H, aromatic); ¹³C-n.m.r. data (CDCl₃): δ 20.47, 20.59 (OAc), 23.31 (NAc), 47.78 (C-2), 61.21 (C-6'), 78.02 (C-3), 97.18 (C-1), and 101.47 (C-1').

Anal. Calc. for $C_{36}H_{45}NO_{15}$: C, 59.09; H, 6.20; N, 1.91. Found: C, 59.16; H, 5.93: N, 2.05.

Benzyl 2-acetamido-6-O-benzyl-2-deoxy-3-O-β-D-galactopyranosyl- α -D-galactopyranoside (14). — A solution of 13 (150 mg) in dry methanol (15 mL) was stirred overnight in the presence of a catalytic amount of the macroreticular¹³ Amberlyst A-26 (OH⁻). The resin was removed by filtration, and the filtrate was evaporated, to give a solid mass that was recrystallized from methanol—ether, to afford 14 in 87% yield (100 mg); m.p. 229–230°, [α]_D +102.5° (c 1.5, Me₂SO); the i.r. spectrum showed the complete absence of ester group.

Anal. Calc. for C₂₈H₃₇NO₁₁: C, 59.67; H, 6.62; N, 2.49. Found: C, 59.59; H, 6.36; N, 2.56.

Benzyl 2-acetamido-6-O-benzyl-2-deoxy-3,4-di-O-(2,3,4,6-tetra-O-acetyl-β-

D-galactopyranoside (15). — Method a. To a stirred solution of 13 (500 mg, 0.68 mmol) and bromide 11 (575 mg, 1.4 mmol) in dry dichloromethane (20 mL) was added 1,1,3,3-tetramethylurea (0.4 mL dissolved in 20 mL of dichloromethane). The flask was then wrapped in aluminum foil, silver triflate (0.36 g) was added, and stirring was continued for 3 days at room temperature. The suspension was filtered through a Celite pad, and the filtrate was successively washed with a saturated solution of sodium hydrogenearbonate and water, dried (anhydrous sodium sulfate), and evaporated. The syrupy product was purified by chromatography on a column of silica gel, with elution with 9:1 (v/v) chloroform–acetone, to give 15 (500 mg, 69%); m.p. 194–195° (chloroform–ether–hexane), $[\alpha]_D$ +33.6° (ϵ 0.6, chloroform); t.l.e. (4:1 chloroform–acetone): R_F 0.36; ¹H-n.m.r. data (CDCl₃): δ 1 90–2.20 (cluster of singlets, 27 H, 8 AcO + NAc) and 7.2–7.3 (m, 10 H, aromatic). + NAc) and 7.2–7.3 (m, 10 H, aromatic).

Anal. Calc. for C₅₀H₆₃NO₂₄; C, 56.54; H, 5.98; N, 1.32. Found: C, 56.79; H, 5.73; N, 1.36.

Method b. In another experiment, diol 8 (401 mg, 1 mmol) was glycosylated exactly as described in (a), using the appropriate quantities of reagents, and the reaction was monitored by t.l.c. in 4:1 chloroform—acetone. After 3 days, t.l.c showed a major spot corresponding to compound 13 and a very minor spot corresponding to compound 15, but also a large amount of unchanged starting-material (8). More bromide 11 (300 mg) and silver triflate (150 mg) were added and stirring was continued for an additional 5 days. T.l.c. then showed that compound 13 was no longer present, but that large amounts of diol 8 still remained. After a total of 8 days, the mixture was processed, and the product purified as described in (a), to give, in 21% yield (220 mg), a pure compound that was identical to compound 15 on the basis of spectral data.

Benzyl 2-acetamido-6-O-benzyl-2-deoxy-3,4-di-O-β-D-galactopyranosyl-α-D-galactopyranoside (16). — O-Deacetylation of compound 15 (300 mg) as described for 14 gave amorphous 16 (180 mg, 88%); [α]_D +76.9° (c 1.1, methanol); the i.r. spectrum showed the complete absence of O-acetyl group; 1 H-n.m.r. data (MeOH- d_4): δ 1.92 (s, 3 H, NAc), 4.87 (d, 1 H, $J_{1.2}$ 4 Hz, H-1), and 7.27 (m, 10 H, aromatic).

Anal. Calc. for $C_{34}H_{47}NO_{16} + H_2O$: C, 54.90; H, 6.64; N, 1.88 Found: C, 54.83; H, 6.71; N, 1.80.

2-Acetamtdo-2-deoxy-3,4-di-O-β-D-galactopyranosyl-D-galactopyranose (17). — A solution of 16 (150 mg) in glacial acetic acid (30 mL) was hydrogenolyzed in the presence of 10% Pd–C (150 mg) for 2 days, the suspension filtered, the filtrate evaporated, and the residue purified by chromatography on a column of silica gel, with elution with 11:9:2 (v/v/v) chloroform-methanol-water, to give amorphous 17 (95 mg, 84%); $[\alpha]_D$ +34.3% (c 1.2, water); t.l.c. in 11:9:2 chloroform-methanol-water: R_F 0.3. The purity of compound 17 was established by paper chromatography on Whatman No. 1 paper with 3.2:1 (v/v) butyl acetate-acetic acid-water, $R_{\rm Gal}$ 0.41 (silver nitrate reagent 14); for 13C-n.m.r. data, see Table II.

Anal. Calc. for $C_{20}H_{35}NO_{16} \cdot H_2O$: C, 42.63; H, 6.62; N, 2.49. Found: C, 42.39; H, 6.45; N, 2.39.

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REFERENCES

- 1 S. S. RANA AND K. L. MATTA, Carbohydr. Res., 113 (1983) c18–c21.
- 2 G. F. SPRINGER, P. R. DESAI, M. S. MURTHY, H. J. YANG, AND E. F. SCANLON, *Transfusion*, 19 (1979) 233-249.
- 3 S. S. RANA, J. J. BARLOW, AND K. L. MATTA, Carbohydr. Res., 84 (1980) 353-357.
- 4 W. D. KLOHS, K. L. MATTA, J. J. BARLOW, AND R. J. BERNACKI, Carbohydr. Res., 89 (1981) 350-354.
- 5 M. M. WEISER, W. D. KLOHS, D. K. PODOLSKY, AND J. R. WILSON, in M. I. HOROWITZ (Ed.), *The Glycoconjugates*, Vol. 4, Academic Press, New York, 1982, pp. 301–334.
- 6 N. V. BOVIN, S. E. ZURABYAN, AND A. YA KHORLIN, Bioorg. Khim., 6 (1980) 789-790.
- 7 H. PAULSEN, Angew. Chem., Int. Ed. Engl., 21 (1982) 155-173
- 8 P. J. GAREGG AND H. HULTBERG, *Carbohydr. Res.*, 93 (1981) C10-C11; P. J. GAREGG, H. HULTBERG, AND S. WALLIN, *ibid.*, 108 (1982) 92-101.
- 9 H. M. FLOWERS AND D. SHAPIRO, J. Org. Chem., 30 (1965) 2041–2043.
- 10 J. GIGG AND R GIGG, J Chem Soc., C, (1966) 82.
- 11 S. HANESSIAN AND J. BANOUB, Carbohydr. Res., 53 (1977) C13-C16.
- 12 J.-C. JACQUINET AND H. PAULSEN, Tetrahedron Lett., (1981) 1387-1390.
- 13 L. A. REED, III, P. A. RISBOOD, AND L. GOODMAN, J. Chem. Soc., Chem. Commun., (1981) 760-761.
- 14 L. HOUGH AND J. K. N. JONES, Methods Carbohydr. Chem., 1 (1962) 21-31.